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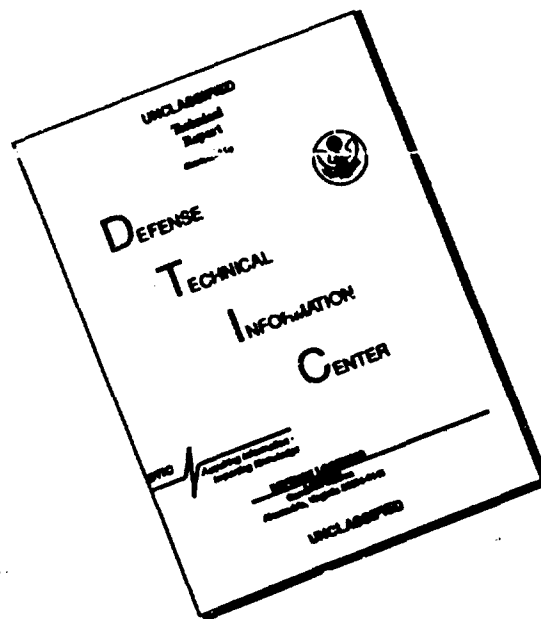
SUPPRESSION OF VIRULENCE  
IN PASTEURELLA PESTIS  
BY THE EPISOME F-LAC

William D. Lawton  
Robert M. Zsigray  
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JANUARY 1967

DEPARTMENT OF THE ARMY  
Fort Detrick  
Frederick, Maryland

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SUPPRESSION OF VIRULENCE IN  
PASTEURELLA PESTIS BY THE EPISOME F-LAC

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Project 1C014501B71A

January 1967

In conducting the research described in this report, the investigators adhered to the "Guide for Laboratory Animal Facilities and Care," as promulgated by the Committee on the Guide for Laboratory Animal Facilities and Care of the Institute of Laboratory Animal Resources, National Academy of Sciences-National Research Council.

#### ABSTRACT

The episome F-lac was transferred from Escherichia coli F-lac into virulent strains of Pasteurella pestis. The resulting strains of P. pestis F-lac were avirulent in mice but segregated (F<sup>-</sup>)lac<sup>-</sup> clones that were fully virulent. This suppression of virulence by F-lac was correlated with a concomitant suppression of the calcium requirement, which is known to be associated with virulence in P. pestis.

## I. INTRODUCTION

The episome F-lac has been transferred between Escherichia coli and species of Salmonella, Shigella, Proteus, Serratia, and avirulent strains of Pasteurella.<sup>1</sup> Evidence is presented here that this episome suppresses the expression of the calcium requirement known to be associated with virulence in P. pestis.

The abbreviations used in this manuscript are: lac, lactose; phe, phenylalanine; cys, cysteine; met, methionine; val, valine; ile, isoleucine; gly, glycine; try, tryptophan; pro, proline; his, histidine.

## II. MATERIALS AND METHODS

### A. STRAINS

Salmonella typhosa try<sup>-</sup> F-lac, S. typhosa try<sup>-</sup> F<sup>2</sup>lac, and S. typhosa try<sup>-</sup> R<sup>+</sup> were obtained from Dr. S. Falkow. Escherichia coli K-12 F-lac strain 23.10.S was obtained from Dr. R. C. Clowes. Other strains used, obtained from our stock collection, were E. coli K-12 pro<sup>-</sup>, Pasteurella pseudotuberculosis type I try<sup>-</sup>, Pasteurella pestis strains Saka, Alexander, and 360, and a methionine-independent histidine-requiring auxotroph from strain Alexander.

### B. MEDIA

Minimal lactose agar medium (designated 1 L) was prepared as follows [final concentration (w/v) in parentheses]. Bacto agar (1.5%) plus 10 ml of a 100 X stock mixture of MgSO<sub>4</sub>·7H<sub>2</sub>O (0.01%), ZnSO<sub>4</sub>·7H<sub>2</sub>O (0.00001%), MnSO<sub>4</sub>·H<sub>2</sub>O (0.00001%), NH<sub>4</sub>Cl (0.053%), and (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> (0.053%) plus 905 ml of distilled water were autoclaved and the following additions made aseptically: 10 ml of 1.0 M potassium phosphate buffer, pH 7.2; 10 ml of a filtered 8 mg/ml stock solution of FeCl<sub>3</sub> (0.008%); 50 ml of 20% lactose (1.0%). This provided 1 liter of medium. Amino acids (usually 0.005%) were added as required. A mixture of six amino acids was necessary for the growth of typical P. pestis: phe (0.01%), cys (0.005%), met (0.005%), val (0.005%), ile (0.005%), and gly (0.005%). This medium is similar to the hemin medium used by Jackson and Burrows.<sup>2</sup>

A complete lactose-indicator medium (BAB-lac) was obtained by supplementing Difco blood agar base with lactose (1%), brom thymol blue (0.0025%), and triphenyl tetrazolium chloride (0.005%). Lac<sup>+</sup> colonies were yellow and lac<sup>-</sup> colonies were red on this medium.

### C. F-LAC TRANSFER

Overnight shake cultures in Difco heart infusion broth (HIB) of donor and recipient strains were usually mixed together for 2 to 4 hours and then plated at appropriate dilutions on a minimal lactose medium formulated to permit only lac<sup>+</sup> recipient cells to form colonies (medium 1 L plus the amino acids required for growth of the recipient). This method was used to pass F-lac from S. typhosa try<sup>-</sup> F-lac into E. coli K-12 pro<sup>-</sup> and subsequently from E. coli K-12 pro<sup>-</sup> F-lac into P. pestis Saka and Alexander. In order to obtain an estimate of F-lac transfer frequency, the method was altered as follows. Approximately 10<sup>8</sup> cells of the recipient lac<sup>-</sup> strain were spread on a minimal lactose agar medium formulated to permit only lac recipient cells to form colonies. The donor F-lac strain was diluted in 0.03 M potassium phosphate buffer, pH 7.2, and a single drop (0.05 ml) of each dilution was spotted on top of the recipient lawn and, as a control, on a sterile plate of the same medium. In this manner, F-lac was transferred from E. coli K-12 F-lac 23.10.S to P. pseudotuberculosis type I try<sup>-</sup> and subsequently from P. pseudotuberculosis type I try<sup>-</sup> F-lac into P. pestis strain 360 (cys<sup>-</sup>phe<sup>-</sup>his<sup>-</sup>).

### D. ASSAY PROCEDURES

Carbohydrate fermentation and urease production were measured by inoculating a suspension of the test organisms into Difco purple broth supplemented with 1% of the desired carbohydrate and into Difco urea broth. Standard methods were used for the detection of virulence,<sup>3</sup> specific antigens,<sup>4</sup> pigmentation,<sup>2</sup> calcium requirement<sup>3</sup> and the production of pesticin, fibrinolytic factor, and coagulase.<sup>5</sup>

## III. RESULTS

The episome F-lac transferred from E. coli K-12 pro<sup>-</sup> F-lac to two virulent strains of P. pestis, Alexander and Saka, at an estimated frequency between 10<sup>-3</sup> and 10<sup>-4</sup> per donor cell. P. pestis F-lac clones were purified and several of the purified clones and their (F<sup>-</sup>)lac<sup>-</sup> spontaneous segregants were tested for virulence in mice (Table 1). Based on ten single lac<sup>+</sup> isolates and nine single lac<sup>-</sup> isolates derived from them, we observed a complete correlation between the presence of F-lac and the phenotypic suppression of virulence. All of the nine lac<sup>-</sup> segregants derived from these phenotypically avirulent F-lac isolates were fully virulent.



These data, plus supporting data obtained with two different virulent strains of *P. pestis*, MP6 and an arginine-requiring auxotroph of Alexander, prompted us to study the correlation in more detail. *P. pestis* strain 360 was infected with F-lac from *P. pseudotuberculosis* type I try<sup>-</sup> F-lac at a frequency of  $10^{-3}$  per donor cell. Fourteen separate lac<sup>+</sup> colonies were purified, tested, and found to be negative for virulence, urease production, and fermentation of rhamnose and melibiose. The latter three properties are characteristically negative in *P. pestis* and positive in *P. pseudotuberculosis*. Satisfied that we could pass F-lac into strain 360 and suppress virulence, we streaked a single colony of strain 360 F-lac five times successively on the selective agar medium and twice on BAB-lac to insure adequate purification. A single lac<sup>+</sup> colony from the final streaking was then streaked on BAB-lac medium and a spontaneous lac<sup>-</sup> segregant clone was picked. A comparison of 360 F-lac, 360 (F<sup>-</sup>)lac<sup>-</sup>, and the original parent strain 360 is presented in Table 2 (virulence) and Table 3 (other characteristics). The suppression of virulence by F-lac was demonstrated as well as the restoration of virulence upon loss of F-lac. Of the 24 properties tested, only the calcium requirement was affected by the presence of F-lac.

TABLE 1. SUPPRESSION OF THE VIRULENCE OF *P. PESTIS* IN MICE BY THE EPISOME F-lac

<i>P. pestis</i> Strain	No. of Individual Colonies Tested	No. Deaths/Total at Dose			
		$2 \times 10^7$	$2 \times 10^5$	$2 \times 10^3$	$2 \times 10^1$
Alexander F-lac	6	2/25	0/30	0/30	0/29
Alexander (F <sup>-</sup> )lac <sup>-</sup>	6	30/30	30/30	30/30	25/30
Saka F-lac	4	0/20	0/20	0/20	0/20
Saka(F <sup>-</sup> )lac <sup>-</sup>	3	15/15	15/15	15/15	14/15

TABLE 2. SUPPRESSION OF THE VIRULENCE OF P. PESTIS STRAIN 360  
IN MICE BY THE EPISOME F-lac

Strain 360 (parent)		Strain 360 F-lac		Strain 360 (F <sup>-</sup> )lac <sup>-</sup>	
Dose	No. Dead/Total	Dose	No. Dead/Total	Dose	No. Dead/Total
358	10/10	$4 \times 10^8$	2/5	27,500	5/5
107	7/10	$4 \times 10^7$	0/5	2,750	4/5
21	5/10	$4 \times 10^6$	0/5	275	4/5
5	1/10	$4 \times 10^5$	0/5	255	4/5
1	0/10	$4 \times 10^4$	0/5	66	4/5
		$4 \times 10^3$	0/5	27	3/5
		$4 \times 10^2$	0/5	14	1/5
				7	0/5

TABLE 3. EFFECT OF THE F-LAC EPISOME ON THE CHARACTERISTICS OF *P. PESTIS*

Property	<i>P. pestis</i> Strains		
	360 (parent)	360 F-lac	360 (F <sup>-</sup> )lac <sup>-</sup>
Production of:			
fraction I antigen	+	+	+
toxin antigen	+	+	+
V antigen	+	+	+
W antigen	+	+	+
pesticin I	+	+	+
fibrinolytic factor	+	+	+
coagulase	+	+	+
pigmentation	+	+	+
urease	-	-	-
Sensitivity to:			
phages T7 and C16	+	+	+
phages T1, T2, T3, T4, T5, T6	-	-	-
Utilization of:			
lactose	-	+	-
melibiose	-	-	-
glycerol	-	-	-
rhamnose	-	-	-
xylose	+	+	+
Growth requirements:			
amino acids	<u>cys,phe,his</u>	<u>cys,phe,his</u>	<u>cys,phe,his</u>
Ca <sup>++</sup> at 37 C	+	-	+

Transfer of F<sup>2</sup>lac or R factor into *P. pestis* strain 360 was accomplished, but the resulting episome-containing isolates of *P. pestis* showed no suppression of virulence or loss of the calcium requirement.

#### IV. DISCUSSION

The suppression of virulence by F-lac should be of interest to plague investigators because it may provide a useful technique to understand virulence better in this pathogen. Our observation supports the opinion of Brubaker and Surgalla<sup>6</sup> concerning the importance of the calcium requirement as a virulence factor. Since the metabolic basis for the calcium requirement is unknown, speculation on possible mechanisms for the suppression of virulence is difficult.

A puzzling aspect of these data is the fact that large numbers of P. pestis F-lac organisms usually did not kill any mice, even though most of the F-lac isolates spontaneously segregated 0.1% to 1.0% lac<sup>-</sup> clones, which are virulent and would be expected to kill the mice in the absence of F-lac cells. Several explanations are possible: (i) the F-lac strains did not segregate in vivo; (ii) the suppression of virulence may be due to the production of a soluble substance that can diffuse into (F<sup>-</sup>)lac<sup>-</sup> cells and suppress virulence; or (iii) the presence of a large excess of F-lac phenotypically avirulent cells may create an immune response in the animals soon enough to prevent the expression of virulence by the (F<sup>-</sup>)lac<sup>-</sup> cells.

From a genetic viewpoint, our observation may lead to further knowledge concerning functions of the F factor. Both F<sup>2</sup>lac<sup>+</sup> and the R factor used in this study are related to the F factor but deficient in their ability to promote chromosomal transfer. Since these two factors did not suppress virulence, it is possible that the suppression of virulence depends on one of the functions of the F factor required for chromosome transfer, such as the ability to attach to the chromosome.

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